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(74) Representative: Hudson, Christopher Mark et **Lilly Industries Limited European Patent Operations Erl Wood Manor** Windlesham Surrey GU20 6PH (GB) Declaration under Rule 28(4) EPC (expert solution)

- (54) Fem A gene of staphylococcus epidermidis, fem A protein, and vectors of microorganisms comprising the fem A gene.
- The instant invention provides the femA gene of Staphylococcus epidermidis and all degenerate sequences thereof, the protein encoded by the femA gene (FemA), and vectors and microorganisms comprising genes encoding the FemA protein.

Clinical isolates of staphylococci (Staphylococcus aureus and S. epidermidis) which cause serious infections due to their intrinsic resistance to beta-lactamase-stable beta-lactam antibiotics (e.g., methicillin) carry the mecA gene. Song et al., FEBS Lett. 221:167-171 (1987). This gene encodes a putative cell wall biosynthetic enzyme referred to as penicillin binding protein 2a (PBP2a). PBP2a, which binds beta-lactams only at concentrations well above therapeutic efficacy, apparently can functionally substitute for all the staphylococcal PBPs and permit growth when the host organism is threatened by beta-lactams. Hartman and Tomasz, J. Bacteriol. 158:513-516 (1984). Wu et al., Antimicrob. Agents Chemother. 36:533-539 (1992) and Ryffel et al., Gene 94:137-138 (1990).

The *mecA* gene is not a normal part of the staphylococcal genome. The organism which donated *mecA* to the staphylococci remains unidentified. Despite the uniform presence of *mecA* in methicillin-resistant clinical isolates, these isolates vary considerably in their degree of resistance to methicillin. This variation in phenotypic expression within a population has been referred to as heterogenous expression. Matthews and Stewart, *FEMS Microbiol. Lett.* 22:161-166 (1984). Typically, most cells exhibit low-level resistance to methicillin and only a minority of the population express high-level resistance, perhaps only one in 10⁸ cells. Tomasz et al., *Antimicrob. Agents Chemother.* 35:124-129 (1991). Although expression of methicillin resistance is dependent upon the presence of FBP2a, it appears to be somewhat independent of the amount of PBP2a, suggesting important roles for other factors. Chambers and Hackbarth, *Antimicrob. Agents Chemother.* 31:1982-1988 (1987) and Murakami and Tomasz, *J. Bacteriol.* 171:874-879 (1989).

Tn551 insertional mutagenesis of methicillin-resistant *S. aureus* revealed numerous sites which influence the level of methicillin resistance but are not linked to *mecA* and do not perturb the expression of PBP2a. Berger-Bächi et al., *Antimicrob. Agents Chemother.* 36:1367-1373 (1992); Kornblum et al., *Eur. J. Clin. Microbiol.* 5:714-718 (1986); Berger-Bächi et al., *Mol. Gen. Genet.* 219:263-269 (1989) and Maidhof et al., *J. Bacteriol.* 173:3507-3513 (1991). Those factors described thus far generally depress the MIC of beta-lactam resistant strains. Some of the genetic loci which demonstrate such an effect on methicillin resistance were designated factors essential for methicillin resistance (*fem*). Berger-Bächi et al., *Mol. Gen. Genet.* 219:263-269 (1989). In contrast to *mecA*, the genes which encode influential factors are probably present in both resistant and susceptible strains of *S. aureus* and *S. epidermidis.* Information obtained from gene disruption studies of *femA* and *femB* in *S. aureus* indicated that in addition to enhanced sensitivity to methicillin, homogeneously methicillin-resistant *S. aureus* strains carrying such gene disruptions have a reduced glycine content in the peptidoglycan component of their cell walls (Maidhof et al., *J. Bacteriol.* 173:3507-3513 (1991)) and exhibit reduced rates of cell wall turnover and autolysis (de Jonge et al., *J. Bacteriol.* 173:1105-1110 (1991)).

Genetic factors, other than *mecA*, that influence the expression of methicillin resistance in *S. epidermidis* have, until now, not been described at the molecular level. The present invention provides DNA sequences encoding the FemA protein of *Staphylococcus epidermidis*, the FemA protein itself, and vectors and microorganisms comprising the *femA* gene of *S. epidermidis*.

The present invention provides DNA sequences encoding the FemA protein of *Staphylococcus epidermi-dis*, including the natural gene sequence designated *femA* (SEQ ID NO:1). Thus, included in the present invention is any DNA compound that comprises an isolated DNA sequence encoding SEQ ID NO:2. SEQ ID NO:2 is as follows:

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	Met 1	Lys	Met	Lys	Phe 5	Thr	Asn	Leu	Thr	Ala 10	Lys	Glu	Phe	Ser	Asp 15	Phe
5	Thr	Asp	Arg	Met 20	Thr	Tyr	Ser	His	Phe 25	Thr	Gln	Met	Glu	Gly 30	Asn	Tyr
10	Glu	Leu	Lys 35	Val	Ala	Glu	Gly	Thr 40	Glu	Ser	His	Leu	Val 45	Gly	Ile	Lys
	Asn	Asn 50	Asp	Asn	Glu	Val	Ile 55	Ala	Ala	Cys	Leu	Leu 60	Thr	Ala	Val	Pro
15	Val 65	Met	ГЛЗ	Ile	Phe	Lys 70	Tyr	Phe	Tyr	Ser	Asn 75	Arg	Gly	Pro	Val	Ile 80
	Asp	Tyr	Asn	Asn	Lys 85	Glu	Leu	Val	His	Phe 90	Phe	Phe	Asn	Glu	Leu 95	Ser
20	Lys	Tyr	Val	Lys 100	Lys	Tyr	Asn	Cys	Leu 105	Tyr	Leu	Arg	Val	Asp 110	Pro	Tyr
	Leu	Pro	Tyr 115	Gln	Tyr	Leu	Asn	His 120	Glu	Gly	Glu	Ile	Thr 125	Gly	Asn	Ala
25	Gly	His 130	Asp	Trp	Ile	Phe	Asp 135	Glu	Leu	Glu	Ser	Leu 140	Gly	Tyr	Lys	His
	Glu 145	Gly	Phe	His	Lys	Gly 150	Phe	Asp	Pro	Va1	Leu 155	Gln	Ile	Arg	Tyr	His 160
30	Ser	Val	Leu	Asn	Leu 165	Ala	Asn	Lys	Ser	Ala 170	Asn	Asp	Val	Leu	Lys 175	Asn
	Met	Asp	Gly	Leu 180	Arg	Lys	Arg	Asn	Thr 185	Lys	Lys	Val	Lys	Lys 190	Asn	Gly
35	Val	Lys	Val 195	Arg	Phe	Leu	Ser	Glu 200	G1u	Glu	Leu	Pro	Ile 205	Phe	Arg	Ser
40	Phe	Met 210	Glu	Asp	Thr	Ser	Glu 215	Thr	Lys	Asp	Phe	Ala 220	Asp	Arg	Glu	Asp
40	Ser 225	Phe	Tyr	Tyr	Asn	Arg 230	Phe	Lys	His	Tyr	Lys 235	Asp	Arg	Val	Leu	Val 240

	Pro	Leu	Ala	Туг	Ile 245	Asn	Phe	Asp	Glu	Tyr 250	Ile	Glu	Glu	Leu	Asn 255	Asn
5	Glu	Arg	Asn	Val 260	Leu	Asn	Lys	Asp	Tyr 265	Asn	Lys	Ala	Leu	Lys 270	Asp	Ile
	Glu	Lys	Arg 275	Pro	Glu	Asn	Lys	Lys 280	Ala	His	Asn	Lys	Lys 285	Glu	Asn	Leu
10	Glu	Gln 290	Gln	Leu	Asp	Ala	Asn 295	Gln	Gln	Lys	Ile	Asn 300	Glu	Ala	Lys	Asn
	Leu 305	Lys	Gln	Glu	His	Gly 310	Asn	Glu	Leu	Pro	Ile 315	Ser	Ala	Gly	Phe	Phe 320
15	Ile	Ile	Asn	Pro	Phe 325	Glu	Val	Val	Tyr	Tyr 330	Ala	Gly	Gly	Thr	Ser 335	Asn
	Arg	Туг	Arg	His 340	Phe	Ala	Gly	Ser	Tyr 345	Ala	Val	Gln	Trp	Lys	Met	Ile
20	Asn	Tyr	Ala 355	Ile	Glu	His	Gly	Ile 360	Asn	Arg	Tyr	Asn	Phe 365	Tyr	Gly	Ile
	Ser	Gly 370	Asp	Phe	Ser	Gļu	Asp 375	Ala	Glu	Asp	Ala	Gly 380	Val	Val	Lys	Phe
25	Lys 385		Gly	Tyr	Asp	Ala 390	Asp	Val	Ile	Glu	Tyr 395	Val	Gly	Asp	Phe	11e 400
30	Lys	Pro	Ile	Asn	Lys 405		Met	Tyr	Asn	Ile 410	Tyr	Arg	Thr	Leu	Lys 415	Lys
	Leu	Lys	Lys													

The natural *femA* sequence is encompassed by the present invention as a DNA compound which comprises the isolated DNA sequence which is SEQ ID NO:1. SEQ ID NO:1 is as follows:

ATG AAG ATG AAG TTT ACG AAT TTG ACA GCT AAA GAA TTT AGT GAC TTT 48 Met Lys Met Lys Phe Thr Asn Leu Thr Ala Lys Glu Phe Ser Asp Phe 15

ACT GAT CGT ATG ACA TAT AGT CAT TTT ACA CAA ATG GAA GGT AAT TAC 96 Thr Asp Arg Met Thr Tyr Ser His Phe Thr Gln Met Glu Gly Asn Tyr 20

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			AAG Lys 35	-													
5			GAT Asp														
10	_		AAA Lys														240
15			AAT Asn														288
			GTA Val														336
20			TAT Tyr 115														384
25			GAT Asp														432
			TTC Phe														480
30			CTA Leu														528
35			GGT Gly														576
40			GTC Val 195														624
	TTT Phe	ATG Met 210	GAG Glu	GAT Asp	ACC Thr	TCT Ser	GAA Glu 215	ACT Thr	AAA Lys	GAT Asp	TTT Phe	GCA Ala 220	GAT Asp	AGA Arg	GAA Glu	GAT Asp	672
45	AGT Ser 225	TTT Phe	TAT Tyr	TAC Tyr	AAC Asn	AGA Arg 230	TTC Phe	AAA Lys	CAT His	TAT Tyr	AAA Lys 235	GAC Asp	CGT Arg	GTT Val	TTA Leu	GTA Val 240	720
50	CCA Pro	CTA Leu	GCC Ala	TAT Tyr	ATT Ile 245	AAC Asn	TTT Phe	GAT Asp	GAG Glu	TAT Tyr 250	ATA Ile	GAG Glu	GAA Glu	CTA Leu	AAT Asn 255	AAT Asn	768

	GAA Glu	AGA Arg	AAT Asn	GTG Val 260	CTT Leu	AAT Asn	AAA Lys	GAT Asp	тат Туг 265	AAT Asn	AAA Lys	GCT Ala	TTA Leu	AAA Lys 270	GAC Asp	ATT Ile	816
5	GAG Glu	AAA Lys	CGT Arg 275	CCA Pro	GAG Glu	AAT Asn	AAA Lys	AAA Lys 280	GCA Ala	CAT His	AAC Asn	AAA Lys	AAG Lys 285	GAA Glu	AAT Asn	TTA Leu	864
10	GAA Glu	CAA Gln 290	CAA Gln	CTC Leu	GAT Asp	GCA Ala	AAT Asn 295	CAG Gln	CAA Gln	AAA Lys	ATT Ile	AAT Asn 300	GAA Glu	GCT Ala	AAA Lys	AAC Asn	912
	TTA Leu 305	AAA Lys	CAA Gln	GAA Glu	CAT His	GGC Gly 310	AAT Asn	GAA Glu	TTA Leu	CCC Pro	ATC Ile 315	TCT Ser	GCT Ala	GGC Gly	TTC Phe	TTT Phe 320	960
15	ATA Ile	ATT Ile	AAT Asn	CCG Pro	TTT Phe 325	GAA Glu	GTA Val	GTT Val	TAC Tyr	TAC Tyr 330	GCT Ala	GGT Gly	GGA Gly	ACT Thr	TCA Ser 335	AAT Asn	1008
20	CGT Arg	TAT Tyr	CGC Arg	CAT His 340	TTT Phe	GCA Ala	GGG Gly	AGC Ser	TAT Tyr 345	GCG Ala	GTT Val	CAA Gln	TGG Trp	AAG Lys 350	ATG Met	ATT Ile	1056
25	AAC Asn	TAT Tyr	GCA Ala 355	Ile	GAA Glu	CAT His	GGT Gly	ATT Ile 360	Asn	CGG Arg	TAT Tyr	AAT Asn	TTC Phe 365	TAT Tyr	GGT Gly	ATT Ile	1104
	AGT Ser	GGT Gly 370	Asp	TTT Phe	AGT Ser	GAA Glu	GAT Asp 375	Ala	GAA Glu	GAT Asp	GCT Ala	GGC Gly 380	GTA Val	GTT Val	AAG Lys	TTT Phe	1152
30	AAA Lys 385	Lys	GGC Gly	TAT	GAT Asp	GCC Ala 390	Asp	GTT Val	ATA Ile	GAA Glu	TAC Tyr 395	Val	GGT	GAC Asp	TTT Phe	ATT Ile 400	1200
35	AAA Lys	CCT Pro	ATT	AAT Asn	AAA Lys 405	Pro	ATG Met	TAT Tyr	AAC Asn	ATT Ile 410	Tyr	AGA Arg	ACA Thr	CTT Leu	AAA Lys 415	Lys	1248
			AAA Lys		7												

Leu Lys Lys

The present invention also includes the protein encoded by SEQ ID NO: 1 in purified form. Also included are recombinant DNA vectors, including expression vectors, that comprise DNA sequences encoding FemA.

The restriction site and function maps presented in the accompanying drawings are approximate representations of the recombinant DNA vectors discussed herein. The restriction site information is not exhaustive; therefore, there may be more restriction sites of a given type on the vector than actually shown on the map.

Figure 1 - A restriction site and function map of plasmid pPSJ180.

Figure 2 - A restriction site and function map of plasmid pET-11A.

The instant invention provides the *femA* gene of *Staphylococcus epidermidis* and all degenerate sequences thereof, the protein encoded by the *femA* gene (FemA), and vectors and microorganisms comprising genes encoding the FemA protein. In the practice of the invention as exemplified herein, the FemA protein comprises the amino acid sequence, which is SEQ ID NO 2:

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	Met 1	Lys	Met	Lys	Phe 5	Thr	Asn	Leu	Thr	Ala 10	Lys	Glu	Phe	Ser	Asp 15	Phe
5	Thr	Asp	Arg	Met 20	Thr	Tyr	Ser	His	Phe 25	Thr	Gln	Met	Glu	Gly 30	Asn	Tyr
40	Glu	Leu	Lys 35	Val	Ala	Glu	Gly	Thr 40	Glu	Ser	His	Leu	Val 45	Gly	Ile	Lys
10	Asn	Asn 50	Asp	Asn	Glu	Val	Ile 55	Ala	Ala	Cys	Leu	Leu 60	Thr	Ala	Val	Pro
15	Val 65	Met	Lys	Ile	Phe	Lys 70	Tyr	Phe	Tyr	Ser	Asn 75	Arg	Gly	Pro	Val	Ile 80
	Asp	Tyr	Asn	Asn	Lys 85	Glu	Leu	Val	His	Phe 90	Phe	Phe	Asn	Glu	Leu 95	Ser
20	Lys	Tyr	Val	L ys 100	Lys	Tyr	Asn	Cys	Leu 105	Tyr	Leu	Arg	Val	Asp 110	Pro	Tyr
	Leu	Pro	Tyr 115	Gln	Tyr	Leu	Asn	His 120	Glu	Gly	Glu	Ile	Thr 125	Gly	Asn	Ala
25	Gly	His 130	Asp	Trp	Ile	Phe	Asp 135	Glu	Leu	Glu	Ser	Leu 140	Gly	Tyr	Lys	His
	Glu 145	Gly	Phe	His	Lys	Gly 150	Phe	Asp	Pro	Val	Leu 155	Gln	Ile	Arg	Tyr	His 160
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	Ser	Val	Leu	Asn	Leu 165	Ala	Asn	Lys	Ser	Ala 170	Asn	Asp	Val	Leu	Lys 175	Asn
5	Met	Asp	Gly	Leu 180	Arg	Lys	Arg	Asn	Thr 185	Lys	Lys	Val	Lys	Lys 190	Asn _.	Gly
	Val	Lys	Val 195	Arg	Phe	Leu	Ser	Glu 200	Glu	Glu	Leu	Pro	11e 205	Phe	Arg	Ser
10	Phe	Met 210	Glu	Asp	Thr	Ser	Glu 215	Thr	Lys	Asp	Phe	Ala 220	Asp	Arg	Glu	Asp
	Ser 225	Phe	Tyr	Tyr	Asn	Arg 230	Phe	Lys	His	Tyr	Lys 235	Asp	Arg	Val	Leu	Val 240
15	Pro	Leu	Ala	Tyr	Ile 245	Asn	Phe	Asp	Glu	Tyr 250	Ile	Glu	Glu	Leu	Asn 255	Asn
20	Glu	Arg	Asn	Val 260	Leu	Asn	Lys	Asp	Tyr 265	Asn	Lys	Ala	Leu	Lys 270	Asp	Ile
20	Glu	Lys	Arg 275	Pro	Glu	Asn	Lys	Lys 280	Ala	His	Asn	Lys	Lys 285	Glu	Asn	Leu
25	Glu	Gln 290	Gln	Leu	Asp	Ala	Asn 295	Gln	Gln	ГЛа	Ile	Asn 300	Glu	Ala	Lys	Asn
	Leu 305	Lys	Gln	Glu	His	Gly 310	Asn	Glu	Leu	Pro	Ile 315	Ser	Ala	Gly	Phe	Phe 320
30	Ile	Ile	Asn	Pro	Phe 325	Glu	Val	Val	туг	Tyr 330	Ala	Gly	Gly	Thr	Ser 335	Asn
	Arg	Tyr	Arg	His 340	Phe	Ala	Gly	Ser	Tyr 345	Ala	Val	Gln	Trp	Lys 350	Met	Ile
35	Asn	Tyr	Ala 355	Ile	Glu	His	Gly	Ile 360	Asn	Arg	Tyr	Asn	Phe 365	Tyr	Gly	Ile
	Ser	Gly 370	Asp	Phe	Ser	Glu	Asp 375	Ala	Glu	Asp	Ala	380	Val	Val	Lys	Phe
40	Lys 385	Lys	Gly	Tyr	Asp	Ala 390	Asp	Val	Ile	Glu	Tyr 395	Val	Gly	Asp	Phe	Ile 400
	Lys	Pro	Ile	Asn	Lys 405	Pro	Met	Tyr	Asn	Ile 410	Tyr	Arg	Thr	Leu	Lys 415	Lys
45	Leu	Lys	Lys									٠				•

The present invention also provides the natural femA gene found in Staphylococcus epidermidis, embodied in SEQ ID NO: 1:

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5								GAC Asp 15	48
J								AAT Asn	96
10								ATT Ile	144
15								GTT Val	192
								GTA Val	240
20				_				TTG Leu 95	288
25								CCA Pro	336
30								AAT Asn	384
								AAA Lys	432
35								TAT Tyr	480
40								AAA Lys 175	528
								AAT Asn	576
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5	GTT Val	AAA Lys	GTC Val 195	Arg	TTT Phe	TTA Leu	TCT Ser	GAA Glu 200	GAA Glu	GAG Glu	TTA Leu	CCT Pro	ATA 11e 205	TTT Phe	AGG Arg	TCA Ser	624
	TTT Phe	ATG Met 210	GAG Glu	GAT Asp	ACC Thr	TCT Ser	GAA Glu 215	ACT Thr	AAA Lys	GAT Asp	TTT Phe	GCA Ala 220	GAT Asp	AGA Arg	GAA Glu	GAT Asp	672
10	AGT Ser 225	TTT Phe	TAT Tyr	TAC Tyr	AAC Asn	AGA Arg 230	TTC Phe	AAA Lys	CAT His	TAT Tyr	AAA Lys 235	GAC Asp	CGT Arg	GTT Val	TTA Leu	GTA Val 240	720
15	CCA Pro	CTA Leu	GCC Ala	TAT Tyr	ATT Ile 245	AAC Asn	TTT Phe	GAT Asp	GAG Glu	TAT Tyr 250	ATA Ile	GAG Glu	GAA Glu	CTA Leu	AAT Asn 255	AAT Asn	768
	GAA Glu	AGA Arg	AAT Asn	GTG Val 260	CTT Leu	AAT Asn	AAA Lys	GAT Asp	TAT Tyr 265	AAT Asn	AAA Lys	GCT Ala	TTA Leu	AAA Lys 270	GAC Asp	ATT Ile	816
20	GAG Glu	AAA Lys	CGT Arg 275	CCA Pro	GAG Glu	AAT Asn	AAA Lys	AAA Lys 280	GCA Ala	CAT His	AAC Asn	AAA Lys	AAG Lys 285	GAA Glu	AAT Asn	TTA Leu	864
25	GAA Glu	CAA Gln 290	CAA Gln	CTC Leu	GAT Asp	GCA Ala	AAT Asn 295	CAG Gln	CAA Gln	AAA Lys	ATT Ile	AAT Asn 300	GAA Glu	GCT Ala	AAA Lys	AAC Asn	912
30	TTA Leu 305	Lys	CAA Gln	GAA Glu	CAT His	GGC Gly 310	AAT Asn	GAA Glu	TTA Leu	CCC Pro	ATC Ile 315	TCT Ser	GCT Ala	GGC	TTC Phe	TTT Phe 320	960
	ATA Ile	ATT Ile	AAT Asn	CCG Pro	TTT Phe 325	Glu	GTA Val	GTT Val	TAC Tyr	TAC Tyr 330	GCT Ala	GGT Gly	GGA Gly	ACT Thr	TCA Ser 335	AAT Asn	1008
35	CGT Arg	тат Туг	CGC Arg	CAT His 340	TTT Phe	GCA Ala	GGG Gly	AGC Ser	TAT Tyr 345	Ala	GTT Val	CAA Gln	TGG Trp	AAG Lys 350	Met	ATT Ile	1056
40	AAC Asn	TAT Tyr	GCA Ala 355	Ile	GAA Glu	CAT	GGT Gly	ATT Ile 360	Asn	CGG Arg	TAT Tyr	AAT Asn	TTC Phe 365	Tyr	GGT Gly	ATT Ile	1104
45	AGT Ser	GGT Gly 370	Asp	TTT Phe	AGT Ser	GAA Glu	GAT Asp 375	Ala	GAA Glu	GAT Asp	GCT Ala	GGC Gly 380	Val	GTT Val	AAG Lys	TTT Phe	1152
45	AAA Lys 385	Lys	GCC	TAT Tyr	GAT Asp	GCC Ala 390	Asp	GTT Val	ATA Ile	GAA Glu	TAC Tyr 395	Val	GGT Gly	GAC Asp	TTT	ATT Ile 400	1200
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	AAA Lys	CC T Pro	ATT Ile	AAT Asn	AAA Lys 405	CCA Pro	ATG Met	TAT Tyr	AAC Asn	ATT Ile 410	TAT Tyr	AGA Arg	ACA Thr	CTT Leu	AAA Lys 415	AAA Lys	1248
	CTA Leu			1257	•										•		

The synthesis of the FemA protein of the present invention may proceed by solid phase peptide synthesis or by recombinant methods. Both methods are described in U.S. Patent No. 4,617,149, the entire teaching of which is herein incorporated by reference. Recombinant methods are preferred if a high yield is desired. The principles of solid phase chemical synthesis of polypeptides are well known in the art and may be found in general texts in the area such as Dugas, H. and Penney, C., *Bioorganic Chemistry* (1981), Springer-Verlag, New York, pgs. 54-92.

Synthesis of the FemA protein can be achieved by recombinant DNA technology. Synthetic genes, the *in vitro* or *in vivo* transcription and translation of which will result in the production of the FemA protein may be constructed by techniques well known in the art. Owing to the natural degeneracy of the genetic code, the skilled artisan will recognize that a sizable yet definite number of DNA sequences may be constructed which encode the FemA protein. All such genes are provided by the present invention. A preferred gene encoding the FemA protein is the natural *femA* gene of *Staphylococcus epidermidis*, which is SEQ ID NO: 1. This preferred *femA* gene is available on an \sim 3.7 kb *EcoRI* restriction fragment of plasmid pPSJ180, publicly available and on deposit in *Escherichia coli* DH5 α at the National Center for Agricultural Utilization Research, 1815 North University Street, Peoria, Illinois 61604-39999, under accession number NRRL B-21024 (date of deposit: December 8, 1992). A restriction site and function map of pPSJ180 is provided in Figure 1 of the drawings.

The femA gene may be created by synthetic methodology. Such methodology of synthetic-gene construction is well known in the art. See Brown et al. (1979) Methods in Enzymology, Academic Press, N.Y., 68:109-151. The femA DNA sequence may be generated using a conventional DNA synthesizing apparatus such as the Applied Biosystems Model 380A or 380B DNA synthesizers (commercially available from Applied Biosystems, Inc., 850 Lincoln Center Drive, Foster City, CA 94404).

To effect the translation of the FemA protein, one inserts the engineered synthetic DNA sequence in any of a large number of appropriate recombinant DNA expression vectors through the use of appropriate restriction endonucleases and DNA ligase. The synthetic *femA* gene should be designed to possess restriction endonuclease cleavage sites at either end of the transcript to facilitate isolation from and integration into these amplification and expression plasmids. The particular endonucleases employed will be dictated by the restriction endonuclease cleavage pattern of the parent expression vector to be employed. The choice of restriction sites are chosen so as to properly orient the FemA coding sequence with control sequences to achieve proper inframe reading and expression of the FemA molecule. The FemA coding sequence must be positioned so as to be in proper reading frame with the promoter and ribosome binding site of the expression vector, both of which are functional in the host cell in which the FemA protein is to be expressed. The FemA protein may be expressed in any number of well-known eucaryotic or procaryotic hosts using known promoters and vectors. Some of the potential hosts, in addition to *E. coli*, include the yeasts *Saccharomyces cerevisiae* and *Pichia pastoris*, *Bacillus*, and cells infected with baculovirus.

To achieve efficient transcription of the synthetic gene, said gene must be operably associated with a promoter operator region. In one practice of the invention, the promoter-operator region of the synthetic gene encoding SEQ ID NO: 2 is placed in the same sequential orientation with respect to the ATG start codon of the synthetic gene as the promoter-operator occupies with respect to the ATG-start codon of the gene from which it was derived. Synthetic or modified promoter operator regions have been created and are well known in the art. When employing such synthetic or modified promoter-operator regions they should be oriented with respect to the ATG-start codon of the *femA* gene as directed by their creators. In one practice of the invention as exemplified herein, where the host cell is an *E. Coli* host cell, said promoter-operator region is the phage T7 promoter-operator region.

A variety of expression vectors useful for transforming procaryotic cells are well known in the art. A preferred vector for expression in an *E. coli* host cell is derived from *E. coli* plasmid pET-11A, which comprises the phage T7 promoter. A restriction site and function map of pET-11A appears in Figure 2 of the accompanying drawings. Plasmid pET-11A is publicly available from Novagen, Inc. (565 Science Drive, Madison, WI 53711) under catalog #69436-1. The preferred host strain is *E. coli* BL21(DE3), also available from Novagen under catalog #69387-1.

The techniques of transforming cells with the aforementioned vectors are well known in the art and may be found in such general references as Sambrook et al., *Molecular Cloning: A Laboratory Manual* (1988), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY or Ausubel et al., *Current Protocols in Molecular Biology* (1989), John Wiley & Sons, New York, NY and supplements. The techniques involved in the transformation of *E. coli* cells used in the preferred practice of the invention as exemplified herein are well known in the art. The precise conditions under which the transformed *E. coli* cells are cultured is dependent on the nature of the *E. coli* host cell line and the expression or cloning vectors employed. For example, vectors which incorporate thermoinducible promoter-operator regions, such as the cl857 thermoinducible lambda-phage promoter-operator region, require a temperature shift from about 30 to about 40°C. in the culture conditions so as to induce

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protein synthesis.

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In a preferred embodiment of the invention *E. coli* K12 BL21 (DE3) cells were employed as host cells but numerous other cell lines are available. The transformed host cells are then plated on appropriate media under the selective pressure of the antibiotic corresponding to the resistance gene present on the expression plasmid. The cultures are then incubated for a time and temperature appropriate to the host cell line employed. Specifically, with *E. coli* K12 BL21 (DE3) cells, the *femA* gene is placed under the control of a promoter transcribed specifically by the T7 RNA polymerase. Induction of transcription of the *femA* gene is accomplished by the addition of isopropylthiogalactoside (IPTG) to the growth medium, which induces expression of the T7 RNA polymerase gene under the control of the *lacUV5* promoter. The T7 RNA polymerase is then available to transcribe the *femA* gene.

General techniques of protein purification are well-known to those of ordinary skill in the art. See Creighton, T.E., Protein Structure: A Practical Approach (1989), IRL Press, Oxford, England and Bollag, D.M. and Edelstein, S.J., Protein Methods (1991), Wiley-Liss, New York, NY. Proteins which are expressed in high-level bacterial expression systems characteristically aggregate in granules or inclusion bodies which contain high levels of the overexpressed protein. Kreuger et al. (1990) in Protein Folding, Gierasch and King, eds., pgs 136-142, American Association for the Advancement of Science Publication No. 89-18S, Washington, D.C. and Sambrook et al., Molecular Cloning: A Laboratory Manual (1988), pp. 17.37-17.41, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY. The FemA protein sometimes aggregates into inclusion bodies when expressed under the control of phage T7 promoter. Such protein aggregates must be solubilized to provide further purification and isolation of the desired protein product. A variety of techniques using strongly denaturing solutions such as guanidinium-HCl and/or weakly denaturing solutions such as dithiothreitol (DTT) are used to solubilize the proteins.

Recombinantly produced proteins may be purified by a variety of techniques well known in the art such as ion exchange chromatography, size exclusion chromatography, electrophoresis, differential centrifugation, reversed phase high performance liquid chromatography, immunoaffinity chromatography, and the like. Protocols for use of these individual techniques or combinations thereof are well known in the art. Gradual removal of the denaturing agents (often by dialysis) in a refolding solution allows the denatured protein to assume its native conformation. The particular conditions for denaturation and refolding are determined by the particular protein expression system and/or protein in question. The S-sulfonates of the peptide molecules are converted to the disulfide paired, folded FemA molecules using a combination of high pH and added thiol in substantial accordance with the teaching of Frank, B.H. et al., (1981) in *Peptides. Synthesis, Structure and Function. Proceedings of the Seventh American Peptide Symposium* (Rich, D.H. and Gross, E., eds.) pp. 729-738, Pierce Chemical Co., Rockford, IL.

The femA gene may be used in gene disruption studies in Staphylococcus epidermidis. Although it is believed that the FemA protein is involved in the formation of a pentaglycine bridge in the cell wall of the bacterium, gene disruption will allow one to ascertain the precise effect of the loss of the femA gene. Gene disruption experiments in Staphylococcus aureus have revealed that a loss of femA results in an ~40% reduction in cell wall glycine content. A similar result might be anticipated for S. epidermidis. Once determined, this information can be used to generate an assay for agents which inhibit the FemA protein, and are therefore useful in combination with antibiotics to treat methicillin-resistant bacteria.

The FemA protein of SEQ ID NO: 2 may be produced by recombinant methods. Recombinant methods are preferred if a high yield is desired. The present invention thus comprises a method for constructing a recombinant host cell capable of expressing SEQ ID NO: 2, said method comprising transforming a host cell with a recombinant DNA vector that comprises an isolated DNA sequence of Claim 1. The present invention also comprises a method for expressing SEQ ID NO: 2 in a recombinant host cell; said method comprising culturing said transformed host cell of Claim 5 under conditions suitable for gene expression.

The following Examples are provided to further illustrate and exemplify, but not limit the scope of, the invention.

Example 1

Source of the Staphylococcus epidermidis femA gene

Isolation of Plasmid pPSJ180

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Alyophil of *E. coli* K12 DH5α/pPSJ180 can be obtained from the Northern Regional Research Laboratories (NRRL), Peoria, Illinois 61604, under the accession number NRRL B-21024 (date of deposit: December 8, 1992). The pPSJ180 plasmid may be isolated from *E. coli* K12 DH5α/pPSJ180 using techniques well-known

to those skilled in the art. See Sambrook et al., Molecular Cloning: A Laboratory Manual (1988), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY or Ausubel et al., Current Protocols in Molecular Biology (1989), John Wiley & Sons, New York, NY and supplements.

5 Isolation of the Staphylococcus epidermidis femA gene via the polymerase chain reaction

Isolated plasmid pPSJ180 is used as the template for the polymerase chain reaction at a concentration of 10 ng/reaction. Vent_R™ DNA polymerase (2 units/µl, Catalog #254, New England Biolabs, 32 Tozer Road, Beverly, MA 01915-9965) is used with standard Vent_R™ DNA polymerase buffer (1X= 10 mM KCl, 20 mM Tris-HCl (pH 8.8 at 25°C), 10 mM (NH₄)₂SO₄, 2 mM MgSO₄, 0.1% Triton X-100). The PCR primers used are AGATA-TAAAGATCTAGATGGGAGTTATGAA (SEQ ID NO: 3) and ATTTCATAATTAGATGGATCCCTTCTTAAAATC (SEQ ID NO: 4). The reaction is carried out by 3 cycles of 94°C for 15 seconds, 40°C for 15 seconds and 72°C for 1 minute followed by 20 cycles of 94°C for 15 seconds, 55°C for 15 seconds and 72°C for 1 minute.

The reaction is transferred to a Centricon 100 microconcentrator (Amicon, Inc., 72 Cherry Hill, Beverly, MA 01915) and washed with 1 ml of water. The microconcentrator is then subjected to centrifugation at 3000 rpm in a microcentrifuge for 30 minutes. The reaction is then diluted to 200 μ l (from \sim 50 μ l) with 1X *Xbal* restriction enzyme buffer. To this is added 50 units of *Xbal* and 50 units of *BamHl*. The DNA is then digested for 90 minutes at 37°C. The DNA is phenol extracted and ethanol precipitated.

20 Example 2

Construction of an Expression Plasmid Containing the femA Gene

The DNA created in Example 1 is then ligated to the 5.6 kb Xbal-BamHI fragment of pET-11A (available from Novagen, Inc. (565 Science Drive, Madison, WI 53711) under catalog #69436-1. This plasmid is then transformed into E. coli BL21 (DE3) (also available from Novagen, Inc. under catalog #69387-1) using techniques well known to those of ordinary skill in the art.

Example 3

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Expression of the FemA Protein

E. coli BL21 (DE3) transformed with the *femA* expression plasmid are grown overnight in TY broth (per liter 10 g tryptone, 5 g yeast extract and 5 g NaCl) and 100 μg/ml ampicillin. The cells are then diluted 1/50 into TY broth + ampicillin and grown at 37°C for 60 minutes. Expression is induced by adding ispropylthiogalactoside (IPTG) to 0.4 mM. Samples are taken at 0 and 6 hours and run on a % SDS-polyacrylamide gel using techniques described in Sambrook et al., *Molecular Cloning: A Laboratory Manual* (1988), pp. 18.47-18.59, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY. An induced protein band is visible by staining with Coomassie Blue at the predicted size of 49,000 daltons.

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SEQUENCE LISTING

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	(1)	GENERA	AL INFORMATION:
10		(i)	APPLICANT: ELI LILLY AND COMPANY (B) STREET: Lilly Corporate Center (C) CITY: Indianapolis (D) STATE: Indiana (E) COUNTRY: United States of America (F) ZIP: 46285
15		(ii)	TITLE OF INVENTION: <u>FEMA</u> GENE OF STAPHYLOCOCCUS EPIDERMIDIS, FEMA PROTEIN, AND VECTORS AND MICROORGANISMS COMPRISING THE <u>FEMA</u> GENE
20		(iii)	NUMBER OF SEQUENCES: 4
20		(iv)	CORRESPONDENCE ADDRESS: (A) ADDRESSEE: C. M. Hudson (B) STREET: Erl Wood Manor (C) CITY: Windlesham
25			(D) STATE: Surrey (E) COUNTRY: United Kingdom (F) ZIP: GU20 6PH
30		(v)	COMPUTER READABLE FORM: (A) MEDIUM TYPE: Floppy disk (B) COMPUTER: Macintosh (C) OPERATING SYSTEM: Macintosh 7.0
35			(D) SOFTWARE: Microsoft Word 5.1
40			
45			

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	(2) INFORMATION FOR SEQ ID NO:1:
5	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1257 base pairs
10	(B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear
15	(ii) MOLECULE TYPE: DNA (genomic)
20	(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 11257
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
30	ATG AAG ATG AAG TTT ACG AAT TTG ACA GCT AAA GAA TTT AGT GAC TTT 48 Met Lys Met Lys Phe Thr Asn Leu Thr Ala Lys Glu Phe Ser Asp Phe 1 5 10 15
35	
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50	

	ACT Thr	GAT Asp	CGT Arg	ATG Met 20	ACA Thr	TAT Tyr	AGT Ser	CAT His	TTT Phe 25	ACA Thr	CAA Gln	ATG Met	GAA Glu	GGT Gly 30	AAT Asn	TAC Tyr	96
5	GAA Glu	TTA Leu	AAG Lys 35	GTT Val	GCT Ala	GAA Glu	GGT Gly	ACC Thr 40	GAG Glu	TCA Ser	CAT His	TTA Leu	GTT Val 45	GGA Gly	ATT Ile	AAA Lys	144
10	AAT Asn	AAT Asn 50	GAT Asp	AAC Asn	GAA Glu	GTG Val	ATT Ile 55	GCA Ala	GCT Ala	TGT Cys	TTA Leu	TTA Leu 60	ACA Thr	GCT Ala	GTT Val	CCT Pro	192
15	GTA Val 65	ATG Met	AAA Lys	ATA Ile	TTT Phe	AAA Lys 70	TAT Tyr	TTT Phe	TAT Tyr	TCC Ser	AAT Asn 75	CGC Arg	GGT Gly	CCA Pro	GTA Val	ATA Ile 80	240
	GAT Asp	TAT Tyr	AAT Asn	AAT Asn	AAA Lys 85	GAG Glu	CTT Leu	GTA Val	CAT His	TTT Phe 90	TTC Phe	TTT Phe	AAT Asn	GAA Glu	TTG Leu 95	AGT Ser	288
20	AAA Lys	TAT Tyr	GTA Val	AAA Lys 100	AAA Lys	TAT Tyr	AAT Asn	TGT Cys	TTA Leu 105	TAT Tyr	TTA Leu	AGA Arg	GTT Val	GAC Asp 110	CCA Pro	TAC Tyr	336
25	CTT Leu	CCA Pro	TAT Tyr 115	CAA Gln	TAT Tyr	TTA Leu	AAT Asn	CAT His 120	GAG Glu	GGA Gly	GAA Glu	ATA Ile	ACT Thr 125	GGA Gly	AAT Asn	GCA Ala	384
30	GGT Gly	CAT His 130	GAT Asp	TGG Trp	ATT Ile	TTT Phe	GAT Asp 135	GAA Glu	TTA Leu	GAG Glu	AGT Ser	TTA Leu 140	GGA Gly	TAT Tyr	AAA Lys	CAC His	432
•	GAA Glu 145	Gly	TTC Phe	CAC His	AAA Lys	GGA Gly 150	TTT Phe	GAT Asp	CCT Pro	GTA Val	TTA Leu 155	CAA Gln	ATC Ile	CGA Arg	TAT Tyr	CAT His 160	480
35	TCT Ser	GTT Val	CTA Leu	AAT Asn	TTA Leu 165	Ala	AAC Asn	AAA Lys	AGT Ser	GCT Ala 170	Asn	GAT Asp	GTT Val	TTA Leu	AAA Lys 175	AAC Asn	528
40	ATG Met	GAT Asp	GGT Gly	TTA Leu 180	AGA Arg	AAG Lys	CGT Arg	AAT Asn	ACT Thr 185	Lys	AAA Lys	GTT Val	AAG Lys	AAA Lys 190	Asn	GGA Gly	576
	GTT Val	AAA Lys	GTC Val 195	Arg	TTT Phe	TTA Leu	TCT Ser	GAA Glu 200	Glu	GAG Glu	TTA Leu	CCT Pro	ATA 11e 205	Phe	AGG Arg	TCA Ser	624
45	TTT Phe	Met 210	Glu	GAT Asp	ACC Thr	TCT	GAA Glu 215	Thr	AAA Lys	GAT Asp	TTT Phe	GCA Ala 220	Asp	AGA Arg	GAA Glu	GAT Asp	672
50	AGT Ser 225	Phe	TAT Tyr	TAC	AAC Asn	AGA Arg 230	Phe	AAA Lys	CAT His	TAT Tyr	AAA Lys 235	Asp	CGT Arg	GTT Val	TTA Leu	GTA Val 240	720

						ATA Ile			768
5						AAA Lys			816
10						AAC Asn			864
15						ATT Ile			912
						ATC Ile 315			960
20						GCT Ala			1008
25						GTT Val			1056
20						TAT Tyr			1104
30						GCT Ala			1152
35						TAC Tyr 395			1200
40						TAT Tyr			1248
		AAA Lvs	1						

(2) INFORMATION FOR SEQ ID NO:2:

5	(i)	SEÇ	QUEN	CE C	HAR	ACTI	ERIS	TIC	s:							
Ü		(2	A) L	ENG	TH:	419	ami	no	acid	is					•	
		(I	3) T	YPE	: am	ino	aci	d								
		(1	o) T	OPOI	LOGY	: 1	inea	ar								
10																
	(ii) MC	OLEC	ULE	TYP	E:]	prot	ein								
15	(xi) SI	EQUE	NCE	DES	CRI	PTIC	n:	SEQ	ID	NO:	2:				
	Met 1	Lys	Met	Lys	Phe 5	Thr	Asn	Leu	Thr	Ala 10	Lys	Glu	Phe	Ser	Asp 15	Phe
20	Thr	Asp	Arg	Met 20	Thr	Tyr	Ser	His	Phe 25	Thr	Gln	Met	Glu	Gly 30	Asn	Tyr
	Glu	Leu	Lys 35	Val	Ala	Glu	Gly	Thr 40	Glu	Ser	His	Leu	Val 45	Gly	Ile	Lys
25	Asn	Asn 50	Asp	Asn	Glu	Val	Ile 55	Ala	Ala	Суз	Leu	Leu 60	Thr	Ala	Val	Pro
	Val 65	Met	Lys	Ile	Phe	Lys 70	Tyr	Phe	Tyr	Ser	Asn 75	Arg	Gly	Pro	Val	Ile 80
30	Asp	Tyr	Asn	Asn	Lys 85	Glu	Leu	Val	His	Phe 90	Phe	Phe	Asn	Glu	Leu 95	Ser
35	Lys	Tyr	Val	Lys 100	Lys	Tyr	Asn	Cys	Leu 105	Tyr	Leu	Arg	Val	Asp 110	Pro	Tyr
	Leu	Pro	Tyr 115	Gln	Tyr	Leu	Asn	His 120	Glu	Gly	Glu	Ile	Thr 125	Gly	Asn	Ala
40	Gly	His 130	Asp	Trp	Ile	Phe	Asp 135		Leu	Glu	Ser	Leu 140	Gly	Tyr	Lys	His
	Glu 1 4 5		Phe	His	Lys	Gly 150	Phe	Asp	Pro	Val	Leu 155	Gln	Ile	Arg	Tyr	His 160
45	Ser	Val	Leu	Asn	Leu 165		Asn	Lys	Ser	Ala 170	Asn	Asp	Val	Leu	Lys 175	Asn
	Met	Asp	Gly	Leu 180		Lys	Arg	Asn	Thr 185		Lys	Val	Lys	Lys 190	Asn	Gly
50	Val	Lys	Val 195	_	Phe	Leu	Ser	Glu 200		Glu	Leu	Pro	Ile 205		Arg	Set

	Phe	Met 210	Glu	Asp	Thr	Ser	Glu 215	Thr	Lys	Asp	Phe	Ala 220	Asp	Arg	Glu	Asp
5	Ser 225	Phe	Tyr	Tyr	Asn	Arg 230	Phe	Lys	His	Tyr	Lys 235	Asp	Arg	Val	Leu	Val 240
40	Pro	Leu	Ala	Туг	Ile 245	Asn	Phe	Asp	Glu	Tyr 250	Ile	Glu	Glu	Leu	Asn 255	Asn
10	Glu	Arg	Asn	Val 260	Leu	Asn	Lys	Asp	Tyr 265	Asn	Lys	Ala	Leu	Lys 270	Asp	Ile
15	Glu	Lys	Arg 275	Pro	Glu	Asn	Lys	Lys 280	Ala	His	Asn	Lys	Lys 285	Glu	Asn	Leu
	Glu	Gln 290	Gln	Leu	Asp	Ala	Asn 295	Gln	Gln	Lys	Ile	Asn 300	Glu	Ala	Lys	Asn
20	Leu 305	Lys	Gln	Glu	His	Gly 310	Asn	Glu	Leu	Pro	Ile 315	Ser	Ala	Gly	Phe	Phe 320
	Ile	Ile	Asn	Pro	Phe 325	Glu	Val	Val	Tyr	Tyr 330	Ala	Gly	Gly	Thr	Ser 335	Asn
25	Arg	Tyr	Arg	His 340	Phe	Ala	Gly	Ser	Tyr 345	Ala	Val	Gln	Trp	Lys 350	Met	Ile
	Asn	Tyr	Ala 355	Ile	Glu	His	Gly	11e 360	Asn	Arg	Туг	Asn	Phe 365	Tyr	Gly	Ile
30	Ser	Gly 370	Asp	Phe	Ser	Glu	Asp 375	Ala	Glu	Asp	Ala	Gly 380	Val	Val	Lys	Phe
	Lys 385	Lys	Gly	Tyr		Ala 390	Asp	Val	Ile	Glu	Tyr 395		Gly	Asp	Phe	Ile 400
35	Lys	Pro	Ile		Lys 405	Pro	Met	Tyr	Asn	Ile 410	Туг	Arg	Thr	Leu	Lys 415	Lys
	Leu	Lys	Lys													

		(2) INFORMATION FOR SEQ ID NO:3:
5		(i) SEQUENCE CHARACTERISTICS:
		(A) LENGTH: 30 base pairs
		(B) TYPE: nucleic acid
40		(C) STRANDEDNESS: single
10		(D) TOPOLOGY: linear
15		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:
		AGATATAAAG ATCTAGATGG GAGTTATGAA 30
20		(2) INFORMATION FOR SEQ ID NO:4:
		(i) SEQUENCE CHARACTERISTICS:
25		(A) LENGTH: 33 base pairs
		(B) TYPE: nucleic acid
		(C) STRANDEDNESS: single
30		(D) TOPOLOGY: linear
35		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:
		ATTTCATAAT TAGATGGATC CCTTCTTAAA ATC 33
40		
	Clai	ms
	1.	A DNA compound that comprises an isolated DNA sequence encoding SEQ ID NO: 2.
45	2.	The DNA compound of Claim 1 which comprises the isolated DNA sequence which is SEQ ID NO: 1.
	3.	A recombinant DNA vector that comprises the isolated DNA sequence of Claim 1.
50	4.	A recombinant DNA vector of Claim 3 that further comprises a promoter positioned to drive expression of said isolated DNA sequence.
	5.	A method for constructing a recombinant host cell capable of expressing SEQ ID NO: 2, said method com-

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transformed host cell of Claim 5 under conditions suitable for gene expression.

7. A recombinant host cell transformed with a recombinant DNA vector of Claim 3.

prising transforming a host cell with a recombinant DNA vector that comprises an isolated DNA sequence

A method for expressing SEQ ID NO: 2 in a recombinant host cell; said method comprising culturing said

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of Claim 1.

8. The protein, in purified form, encoded by SEQ ID NO:2.

FIG. I

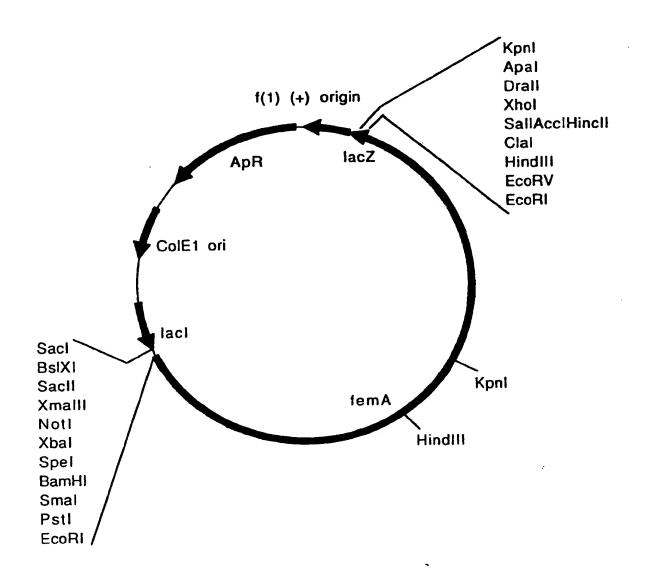
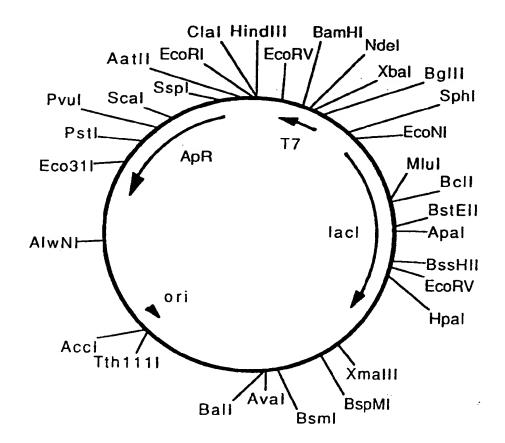


FIG. 2



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Declaration under Rule 28(4) EPC (expert solution)

- Fem A gene of staphylococcus epidermidis, fem A protein, and vectors of microorganisms comprising the fem A gene.
- The instant invention provides the femA gene of Staphylococcus epidermidis and all degenerate sequences thereof, the protein encoded by the femA gene (FemA), and vectors and microorganisms comprising genes encoding the FemA protein.

EP 0 625 575 A3



EUROPEAN SEARCH REPORT

Application Number EP 94 30 2950

	DOCUMENTS CONSI	DERED TO BE RELEVAN	T	
ategory	Citation of document with it of relevant pa	ndication, where appropriate,	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.CL5)
•	MOL. GEN. GENET., vol.219, 1989 pages 263 - 269 BERGER-BÄCHI, B. ET host-mediated factomethicillin resista aureus: Molecular characterization'see Table 1 and Fig.	r essential for nce in Staphylococcus loning and	1-8	C12N15/31 C12P21/02 C07K13/00
Y	vol.36, no.12, Dece pages 2617 - 2621 HURLIMANN-DALEL, R.	ET AL. 'Survey of the nce-associated genes and femA-femB in of methicillin	1-8	
A	WO-A-91 08305 (U-GE 1991 see pages 1-4	NE RESEARCH) 13 June	1	TECHNICAL FIELDS SEARCHED (Int.CL.5) CO7K
	The present search report has	been drawn up for all claims		
	Place of search	Date of completion of the search		Examiner
Y:pan do A:tex O:no	MUNICH CATEGORY OF CITED DOCUME recurrency of taken alone reticularly relevant if taken alone reticularly relevant if conshined with an current of the same category shoological background as-written disclosure ermodiate document	E : earlier patent of after the filing	iple underlying the locument, but pul- date d in the application of for other reasons	obished on, or

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